

Case Studies: Introduction to Cardiovascular & Systemic Diseases

Systemic infections can be caused by many different infectious agents: bacterial, fungal, viral, and parasitic. One common finding for all systemic infections is the need for a **portal of entry**. The portal of entry can be via the parenteral route (as in mosquito-borne diseases such as malaria), via the oral route (as in typhoid fever), via sexual contact (as in HIV infection), as a bloodborne pathogen (as in hepatitis B virus infection), via the respiratory tract (as in measles), and by horizontal transmission via transplacental infection (as in congenital cytomegalovirus infection). In many cases of systemic infection, **colonization** occurs prior to the dissemination of the infectious agent throughout the body. In some diseases (e.g., tetanus and diphtheria) the infection itself is caused by a noninvasive organism and the systemic symptoms are caused by the dissemination of a **toxin** that is responsible for the disease. In most cases, however, the etiologic agent is disseminated via the hematogenous route.

Patients may have certain risk factors or defects in host defenses that predispose them to specific types of infections. Examples of defects in host defenses that predispose to certain specific types of infections include breaches in the integrity of the skin (patients with burns, patients with invasive medical devices), defects in cell-mediated immunity (AIDS, corticosteroids), defects in humoral immunity (hypogammaglobulinemia), decreased splenic function (splenectomy, sickle cell disease), quantitative defects in neutrophils (neutropenia following chemotherapy), qualitative defects in neutrophils (chronic granulomatous disease, Chediak-Higashi syndrome), and deficiencies in the complement system. It is important to be able to recognize these risk factors when they are present and to know to what the defect predisposes the patient. Conversely, it is important to be able to suspect a specific defect in host defenses when a patient presents with a systemic infection.

Protection of the host from a systemic infection can occur as a result of a prior infection with the specific agent of infection (e.g., measles) or due to a vaccination to that agent. Unfortunately, efficacious vaccines are not available for the majority of infectious agents, and in many diseases, infection does not lead to protective immunity. Important agents of systemic infection are listed in Table 6. Please note that virtually all bacteria can potentially be isolated from the blood under circumstances of specific host defects, such as the presence of an intravenous catheter. Many of the etiologic agents listed have a particular organ tropism (such as the liver for hepatitis viruses) but may cause systemic illness.

Organism	General Characteristics	Source of Infection	Disease Manifestation
Bacteria			
<i>Acinetobacter</i> spp.	Lactose-nonfermenting, gram-negative bacilli	Exogenous	Nosocomial UTI, nosocomial pneumonia, nosocomial and line-related bacteremia
<i>Bartonella henselae</i>	Fastidious, gram-negative bacillus	Exogenous; cats appear to be primary host	Cat scratch disease; bacillary angiomatosis (in immunocompromised)
<i>Borrelia burgdorferi</i>	Spirochete	Tick borne	Lyme disease; rash, arthritis, nervous system and cardiac manifestations
<i>Brucella</i> spp.	Oxidase-positive, gram-negative bacilli	Zoonosis	Lymphadenopathy, hepatosplenomegaly; genitourinary, bone, and CNS infection
<i>Clostridium botulinum</i>	Anaerobic gram-positive bacillus	Exogenous	Botulism
<i>Clostridium perfringens</i>	Anaerobic gram-positive bacillus	Exogenous	Gas gangrene, emphysematous cholecystitis, bacteremia, food poisoning
<i>Clostridium tetan</i>	Anaerobic gram-positive bacillus	Exogenous	Tetanus

Organism	General Characteristics	Source of Infection	Disease Manifestation
Coagulase-negative staphylococci	Catalase-positive, coagulase-negative, gram-positive cocci	Endogenous	Nosocomial bacteremia
<i>Corynebacterium diphtheriae</i>	Aerobic gram-positive bacillus	Exogenous	Diphtheria
<i>Enterobacter</i> spp.	Lactose-fermenting, gram-negative bacilli	Endogenous	Community and nosocomial UTI, bacteremia, intra-abdominal infections
<i>Enterococcus</i> spp.	Catalase-negative, gram-positive cocci	Endogenous	Wound infections, nosocomial UTI, bacteremia, endocarditis
<i>Escherichia coli</i>	Lactose-fermenting, gram-negative bacillus	Endogenous	Community and nosocomial UTI, bacteremia, intra-abdominal infections
<i>Francisella tularensis</i>	Gram-negative bacillus	Zoonosis	Ocular, lymphadenopathy, pulmonary, bacteremia
Group A streptococci (<i>Streptococcus pyogenes</i>)	Catalase-negative, gram-positive cocci	Endogenous	Pharyngitis, cellulitis, bacteremia, scarlet fever, necrotizing fasciitis, pneumonia, poststreptococcal glomerulonephritis and rheumatic fever
Group B streptococci (<i>Streptococcus agalactiae</i>)	Catalase-negative, gram-positive cocci	Endogenous	Sepsis, meningitis, cellulitis
<i>Klebsiella pneumoniae</i>	Lactose-fermenting, gram-negative bacillus	Endogenous	Community and nosocomial UTI, bacteremia, intra-abdominal infections
<i>Mycobacterium avium</i> complex	Acid-fast bacilli	Exogenous	Disseminated disease
<i>Mycobacterium tuberculosis</i>	Acid-fast bacillus	Respiratory, may be exogenous (primary) or endogenous (reactivation)	Pneumonia, extrapulmonary tuberculosis, miliary tuberculosis
<i>Neisseria meningitidis</i>	Oxidase-positive, gram-negative diplococcus	Endogenous (from colonization)	Meningitis, bacteremia
<i>Pasteurella multocida</i>	Oxidase-positive, gram-negative bacillus	Zoonosis (often animal bite or scratch)	Cellulitis, bacteremia, osteomyelitis, meningitis
<i>Proteus mirabilis</i>	Lactose-nonfermenting, gram-negative bacillus	Endogenous	Community and nosocomial UTI, bacteremia
<i>Pseudomonas aeruginosa</i>	Lactose-nonfermenting, Oxidase-positive, gram-negative bacillus	Exogenous	Community and nosocomial UTI, nosocomial pneumonia, nosocomial bacteremia
<i>Rickettsia prowazekii</i>	Rickettsial organism	Exogenous, lice to human	Epidemic typhus causes fever and disseminated intravascular coagulation.

Organism	General Characteristics	Source of Infection	Disease Manifestation
<i>Rickettsia rickettsii</i>	Rickettsial organism	Exogenous, tick to human	Rocky Mountain spotted fever
<i>Salmonella typhi</i>	Lactose-nonfermenting, gram-negative bacillus	Exogenous, human to human	Typhoid fever, bacteremia, intestinal disease
<i>Staphylococcus aureus</i>	Catalase-positive, coagulase-positive, gram-positive coccus	Endogenous	Skin infections, bacteremia, endocarditis, septic arthritis, abscesses
<i>Streptococcus pneumoniae</i>	Catalase-negative, gram-positive coccus	Endogenous	Community-acquired pneumonia, sinusitis, meningitis, bacteremia, endocarditis
<i>Treponema pallidum</i>	Spirochete (does not Gram stain)	Direct sexual contact, vertical (mother to child)	Primary, secondary, latent, and late syphilis; can affect any organ
<i>Viridans group streptococci</i>	Catalase-negative, gram-positive cocci	Endogenous	Endocarditis
<i>Yersinia pestis</i>	Gram-negative bacillus	Zoonosis; person to person in pneumonic form	Lymphadenopathy (bubonic), pneumonia, bacteremia

Fungi

<i>Aspergillus</i> spp.	Molds with septate hyphae	Exogenous	Pneumonia, sinusitis, external otitis, allergic processes, disseminated infection
<i>Blastomyces dermatitidis</i>	Dimorphic mold	Exogenous	Pneumonia, meningitis, bone infection
<i>Candida albicans</i>	Yeast, often germ tube positive	Endogenous	Thrush, vaginal yeast infection, diaper rash, esophagitis, nosocomial UTI, nosocomial bloodstream infection
<i>Candida</i> spp., non-albicans	Yeasts, germ tube negative	Endogenous	Thrush, vaginal yeast infection, nosocomial UTI, nosocomial bloodstream infection
<i>Coccidioides immitis</i>	Dimorphic mold	Exogenous	Pneumonia, meningitis, bone infection
<i>Cryptococcus neoformans</i>	Encapsulated yeast	Exogenous	Meningitis, pneumonia, bloodstream infection
<i>Histoplasma capsulatum</i>	Dimorphic mold	Exogenous	Pneumonia, disseminated infection
<i>Zygomycetes</i>	Molds with aseptate hyphae	Exogenous	Pneumonia, sinusitis, invasive infection

Viruses			
Dengue Virus	Enveloped polyhedral capsid with single-stranded RNA	Exogenous (<i>Aedes aegypti</i> mosquito)	Dengue Fever; joint pain, fever, headache, rash, muscle pain, hemorrhagic fever/shock
Filoviruses	Enveloped helical capsid with single-stranded RNA	Exogenous: unknown possible animal reservoir	Hemorrhagic fever with high mortality.
Parasites			
<i>Babesia microti</i>	Can be seen on peripheral blood smear	Exogenous (ticks)	Babesiosis
<i>Leishmania donovani</i>	Amastigotes in tissue touch preparation	Exogenous (<i>Phlebotomus</i> fly)	Kalaazar
<i>Plasmodium</i> spp.	Can be seen on peripheral blood smear	Exogenous (<i>Anopheles</i> mosquito)	Malaria
<i>Strongyloides stercoralis</i>	Nematode	Exogenous; endogenous (autoinfection and hyperinfection)	Gastrointestinal, pulmonary (pneumonia, wheezing), disseminated in hyperinfection
<i>Taenia solium</i>	Tapeworm	Exogenous	Gastrointestinal infection, cysticercosis (brain, muscles, other organs)
<i>Toxoplasma gondii</i>	Protozoan	Exogenous; endogenous (reactivation)	Central nervous system, ocular, hepatic, pulmonary

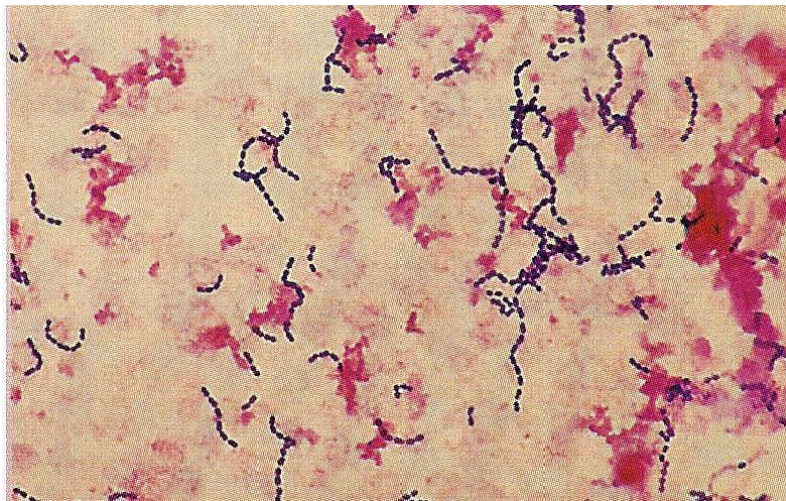
Case One

This 39-year-old intravenous drug user (actively using cocaine on the date of admission) was admitted with cellulitis of the right arm after experiencing fevers for several weeks. He had been treated with outpatient antibiotics without relief of either associated chills or dizziness. Two sets of blood cultures were obtained on admission. A trans-thoracic echocardiogram demonstrated a 1cm vegetation on the ventral surface of the aortic valve. The patient left the hospital against medical advice, but was readmitted 2 days later for antimicrobial therapy.

Past medical history was notable for multiple hospital admissions for both cellulitis and abscesses primarily involving the patient's arm. He had had multiple drug rehabilitation treatment attempts without success.

Physical examination demonstrated a thin, unkempt man in no acute distress with multiple "needle track" marks on both his upper and lower extremities. No splinter hemorrhages or signs of embolic phenomena were noted on the extremities. Cardiac exam was notable for a grade II/VI systolic murmur best heard at the left sternal border. The spleen tip was palpable. The right arm had a 10-by-6-cm excoriated area with surrounding induration.

Gram stain of an organism detected in both sets of the blood cultures obtained at admission. The organism grew in broth containing 6.5% NaCl, hydrolyzed esculin in the presence of bile (i.e., was bile esculin positive), and was catalase



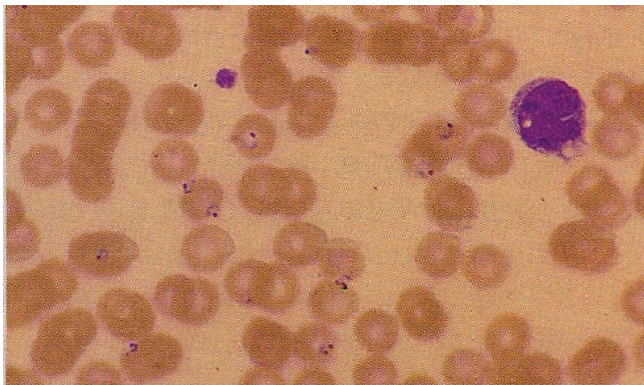
negative.

1. What type of infection does this patient have?
2. What organisms frequently cause this type of infection in intravenous drug users (IVDUs)? What organism is causing his infection?
3. How does intravenous drug use predispose the patient to this type of infections? Briefly describe the pathogenesis of this infection. Describe what other organs may be secondarily infected and the mechanism by which secondary infections occur.
4. When considering antimicrobial therapy for this infection, what general strategy should be employed?
5. What antimicrobial resistance problems have recently emerged involving this organism? What strategies have been employed to reduce the spread of these organisms?

Case Two

The patient was a 41-year-old man who had returned from Central Africa 12 days prior to his admission. He had been in Africa for approximately 7 months, working as a civil engineer on road building projects in Rwanda, Zaire, and Uganda. Seven days before admission, the patient noted the acute onset of fevers with chills as well as cough and myalgias. For the next 3 days he had episodes of fevers and chills approximately three times a day. After 3 days of fever, he sought care at a local urgent care center. He told the physician that he had just returned from Africa and had not taken malaria prophylaxis. He had a white blood cell count of $2,200/\mu\text{l}$ and a platelet count of $102,000/\mu\text{l}$. No hematocrit was reported. A PPD test was placed. The patient was diagnosed as having bronchitis, treated with azithromycin, and instructed to return in 48 h to have his PPD read.

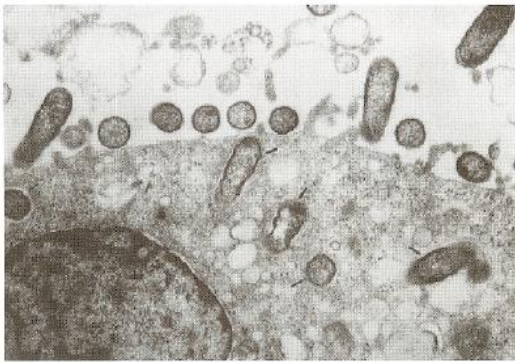
On his return to have his PPD read, the patient was found to be hypotensive and had mental status changes. He was noted to be icteric. He was referred to the emergency room of the local hospital to "rule out hepatitis." A diagnosis was made in the hematology laboratory (blood stain below). At the time of his diagnosis, he was noted to have a hematocrit of 25%. After 24 h of antimicrobial and aggressive fluid therapy, he had a cardiopulmonary arrest. He was resuscitated. In the next 24 h, he developed acute respiratory distress syndrome and became progressively hypotensive. He was transferred to our hospital. On arrival, he was intubated and comatose. On physical examination, he was grossly icteric and tachycardic and was noted to be oozing bright red blood from his mouth and catheter sites. His laboratory tests were significant for a hematocrit of 23%, platelets of $39,000/\mu\text{l}$, prolonged bleeding times, and 3+ hemoglobin and 25 to 50 red blood cells in his urine. He was given fresh frozen plasma, platelets, and cryoprecipitate for his disseminated intravascular coagulation. Exchange transfusion was begun, with the patient receiving seven units of packed red blood cells. His condition continued to deteriorate, and he had a cardiac arrest from which he could not be resuscitated.



1. With what organism was this patient infected? (You should be able to identify this organism to the species level.)
2. Describe two complications associated with this organism, including the pertinent clinical findings seen in this case. What unique characteristic of this organism is important in producing these complications, and how did it contribute to producing these complications?
3. Why do you think exchange transfusion was used as a therapeutic strategy in this patient?
4. Discuss the problems associated with chemotherapy for infection with this organism.

Case Three

An extensive ongoing outbreak of typhus occurred in refugee camps in Rwanda, Burundi, and Zaire. For people in the camps, daily living was an immense hardship. Following the outbreak of civil war in 1993, over 760,000 refugees lived in camps under appalling conditions. Sanitation and clean water were hard to find, and disease was rampant. Besides typhus, outbreaks of typhoid fever, dysentery, and malaria also affected the refugee communities. The United Nations World Food Program distributed emergency rations to curb malnutrition. In some refugee camps, the people were required to work like a chain gang, collectively, on a single tract of land at a time. Those that left the camps were assumed to be rebel forces and could be shot by government soldiers. The civil war between the two main tribes for control of the government degraded into a tit-for-tat massacre of civilians. Against this background, a typhus epidemic emerged among the displaced population of Burundi. The outbreak may have begun among prisoners in a jail in N'Gozi in 1995. Clinical aspects of the disease included headache, chills, fever, prostration, confusion, photophobia, vomiting, and rash (seen below, generally starting on the trunk). There was a fatality rate of 15% among jail inmates. At the time, the disease was not recognized and was referred to as sutama. Reports of sutama among the civilian population date back to late 1995, and in association with body louse infestation, the disease subsequently swept across the higher and colder regions of the country. During a field study in February 1997, 102 refugees with sutama underwent clinical examinations and interviews. Serum samples were collected, and infesting body lice (pictured below) were removed. Analysis of blood sera by immunofluorescence microscopy found antibodies present against the causative agent, a small obligately intracellular pathogen (TEM below). Most of the 102 patients with sutama during initial assessment presented with typical manifestations of the disease. Up to September 1997, 45,558 cases were clinically diagnosed, most of which occurred in regions at an altitude of over 1,500 meters.



1. What pathogen caused the typhus outbreak?
2. How is the pathogen typically transmitted?
3. How does the pathogen cause the bleeding under the skin that leads to the rash?
4. What recommendations would you have made to treat this pathogen? Explain why.
5. Why would the disease be most common in higher altitudes of eastern Africa?
6. Why did the outbreak first appear among prisoners?

Case Four

A 24-year old, female graduate student in biology presented with exhaustion, weakness and a low grade fever. She was pale and showed poor ability to concentrate. Her history revealed that she had gradually become increasingly tired and weakened over the past two months. She had experienced low-grade fevers over the past month and felt she would need to drop out of her graduate program if she did not get this under control. She had had a severe strep throat about a year earlier and showed some signs of rheumatic fever at the time. She had had minor dental surgery about two months earlier.

On examination, she had a temperature of 100°F. She had slightly enlarged cervical lymph nodes. She had a heart murmur, with abnormal valve sounds. Her ears, eyes and throat were clear. She had clear lungs and there were no significant findings in other systems. Organisms were isolated from her blood and plated on blood agar (Figure 1).

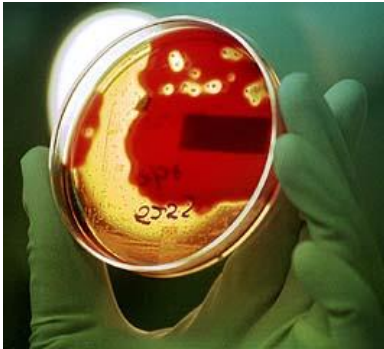


Figure 1

1. What would be your primary diagnosis of this patient?
2. What agent do you think is causing this problem?
3. How would you make a definite diagnosis?
4. What are the possible outcomes when this disease is left untreated?